## Attachment 2

## 2003 Tuolumne River Fall Chinook Salmon Escapement Survey

Prepared by:
Dennis Blakeman
Fisheries Biologist California Department of Fish and Game

March 2004

## INTRODUCTION

The San Joaquin fall-run Chinook salmon is currently a candidate species under the Federal and State Endangered Species Acts. Population levels in the Tuolumne River have declined in the latter half of the $20^{\text {th }}$ century from a high of approximately 130,000 returning adults in 1944 (Fry 1961) to a low of 77 in 1991 (Neillands et al. 1993). Current levels of 7,916 in 1998 (Heyne 1998), 7,685 in 1999 (Heyne 2000), 17,873 in 2000 (Vasques 2001), 9,222 in 2001 and 7,125 in 2002, indicate a slight recovery period. The decline of the species is believed to be caused by many factors. In general, reduction of spawning and rearing habitat and stream flow management practices are thought to be the major factors limiting overall population numbers. Numerous additional factors including but not limited to predation, streambed alteration, pump diversion, gravel mining, land use practices, and ocean angler harvest contribute to a web of complex population dynamics which effect population numbers within the habitat currently available to Tuolumne River Chinook salmon.

The California Department of Fish and Game (CDFG) has conducted escapement surveys on the Tuolumne River since 1940 (Fry 1961). The Schaefer mark recapture escapement estimation model (Schaefer 1951) has been utilized since 1971. The 2003 escapement survey will begin using the JollySeber (Seber 1973) escapement model but will continue to report Schaefer estimates. Beginning in 1992, CDFG escapement surveys have been utilized as part of the New Don Pedro FERC Project No. 2299 license monitoring program and annual reporting.

The primary objectives of the Tuolumne River escapement survey are to:

- Estimate the escapement of fall run Chinook salmon on the Tuolumne River.
- Collect fork length and sex data.
- Collect scale and otolith samples with which to conduct age determination analysis and subsequent cohort analysis.
- Collect and analyze coded wire tag data from marked hatchery fish.
- Evaluate the distribution of salmon redds through the study area.
- Collect DNA samples for storage at the CDFG Salmonid Tissue Archive for subsequent analysis.


## STUDY AREA

Approximately 26.5 river miles were surveyed during the Tuolumne River escapement survey in 2003 (Figure 1). The survey area was divided into 4 sections with Section 1 being the upstream most reach. Section 1, also referred to as the primary spawning reach, extends from riffle A1 at river mile 51.6 near La Grange Dam downstream to Basso Bridge at river mile 47.5. Section 2 extends from Basso Bridge down to the Turlock Lake State Recreation Area (TLSRA) at river mile 41.9. Section 3 covers the area between TLSRA and riffle S1 at river mile 34. Section 4 extends downstream to Fox Grove (river mile 26).

All riffles in the study area have been identified and mapped using a Trimble GPS unit and the GIS computer program ArcView. Each riffle has been systematically re-named upstream to downstream using sequential letter/number designations for river mile and riffle number, respectively. For example, the first riffle immediately below La Grange Dam in the first river mile (56) is named A1. This numbering system is a departure from the historical riffle numbering system. However, the new riffle identification system is more logical and is more conducive to editing as river morphology changes. The riffle identification cross-reference is located in Table 1.

## METHODS

## Population Estimation

The Schaefer (1951) and Jolly-Seber (Seber 1972) mark recapture models were used to estimate fall salmon escapement on the lower Tuolumne River. These methods utilize marked and subsequently recovered carcasses during weekly surveys of the spawning reach. A ratio of marked to unmarked fish is used to calculate weekly population estimates, which are then summed to estimate the total spawning population. The CDFG began the survey on 30 September 2003 (Week 1) and concluded on 6 January 2004 (Week 15). Carcasses were tagged for the first 13 weeks. Weeks 14 and 15 no carcasses were tagged, these were strictly carcass recovery weeks. During the two recovery weeks, carcasses were collected and examined for jaw tags and all carcasses collected were chopped in half.

All carcasses encountered were handled during weekly drift boat surveys of the study area. Carcasses were gaffed as the sampling crew drifted past and held in the boat until the end of the riffle and adjacent downstream pool. Subsequent to drifting the riffle and downstream pool the riverbanks were walked to collect carcasses that could not be seen or collected from the drift boat. Every carcass handled was
designated as fresh, decayed, skeleton or recovery, depending on the degree of decomposition or the presence of an aluminum jaw tag in the case of recoveries. The fresh carcass designation criteria during 2003 was at least one clear eye (Figure 2). Decayed fish had cloudy eyes. Skeletons were carcasses judged to be in an advanced state of decay and unlikely to have the same probability of recapture as fresh and decayed specimens. Criteria for skeleton designation during the 2003 survey included the presence of fungus covering the entire body at the freshest end of skeleton designation (dead approximately one week) to actual skeletons at the most decayed end (Figures 3 and 4).

All fresh and decayed carcasses were given a unique number by attaching a numbered aluminum tag to the lower jaw. These newly tagged carcasses were redistributed to river current near the lower end of the riffle for recovery in subsequent weeks. For tag recoveries, the unique tag number was noted and the carcass was chopped and returned to the river. All skeletons were enumerated, chopped, and returned to the river to avoid double counting despite findings by Law (1994) suggesting that untagged carcasses not removed after initial count only slightly affected Schaefer's (1951) population estimate. Estimates were made using the Schaefer (1951) equation as presented in Ricker (1975) and also using the Jolly-Seber equation (Seber 1973). Law (1994) found in simulations of various models, using a similar protocol as this survey, that the Peterson model (see Ricker, 1975) drastically over estimated, while the Schaefer model consistently overestimated the population and the Jolly-Seber model most accurately estimated the population. Therefore, Peterson's model was not used in this analysis and the Jolly-Seber model will now be included with Schaefer estimates.

## Weekly Fish Distribution and Redd Counts

Weekly live fish observation and redd counts were conducted during the survey (Table 2, Figure 5). These counts are conducted for each riffle and pool using the riffle identification system noted earlier. Counts are made using tally counters as field crews drifted through riffles and pools.

## Individual Fish Data Collection

Fork length (to the nearest 1 centimeter) and sex data are collected for all tagged carcasses. Scale and otolith samples are collected from a percentage of specimens to determine the size and age composition of annual spawning runs. Coded wire tags (CWTs) are collected from hatchery produced, marked (adipose fin clipped), carcasses as part of long term survival testing of releases of marked outmigrating smolts. This also allows for determining the incidence of straying from other river systems. CWT specimens are also used to validate scale and otolith age determination work. Genetic samples: caudal, dorsal, or pectoral fin clips were collected, and delivered to the CDFG Salmonid Tissue Archive at the end of the
survey. Scale and otolith samples were collected from both wild and CWT carcasses and are catalogued at the CDFG La Grange Field Office. CWTs and otoliths are collected via removal of the head minus the lower jaw. Extraction and analysis of otoliths and CWTs is conducted after the spawning season. All fish samples are catalogued by the fish's unique jaw tag number, which allows the samples to be tracked to the specific data and riffle number of collection.

## RESULTS

## Population Estimate

Based on the Jolly-Seber model using all fish the 2003 escapement estimate was $\mathbf{2 , 1 6 3}$ salmon. The Jolly-Seber model using all tagged fish and recoveries yields the most accurate estimate. The Schaefer model utilize the number of recoveries of tagged carcasses that were fresh when tagged, the total number of fresh tagged fish, and the total number carcasses handled each week to generate weekly escapement estimates (Table 3). Weekly estimates are summated to estimate total escapement over the course of the survey. Table 4 shows the total number of fresh tagged each week in relation to the number of recoveries made in subsequent weeks. Weekly estimates are presented in Table 5. The Jolly-Seber calculation matrix required that tagging and recapture numbers be shifted to reflect a continuous recovery period. Thus, the one recovery in week three was moved to week five, and for calculation purposes recovery week five became recovery week two (Table 4-5). Weekly cumulative Schaefer and Jolly-Seber estimates are graphed in Figure 6. The fresh tagged recovery rate was $55.0 \%$ which is slightly lower than the overall recovery rate of $56.8 \%$ and the overall recovery rates of $64.4 \%$ in 2002 and $61.3 \%$ encountered during the 2001 escapement survey.

## Weekly Counts

Live fish counts increased steadily, peaked in weeks 7 and 8 , and declined steadily through the remainder of the survey (Table 2, Figure 5). Carcass counts exhibited a similar incline, peak, and decline which were offset from live counts by about one week. The carcass count peaked in weeks 8 and 9 . Redd counts increased through Week 8 when the total number of observations was 349.

## Spawning Distribution

The results of total weekly redd counts clearly indicate that the majority of spawning activity is concentrated in the riffles of Section 1 (Figures 7 and 8). The maximum number of redds counted in a particular riffle over the course of the season are listed in Table 6. The maximum redd count represents the redd count made when external factors like visibility were at optimum conditions. During the 2003
survey 649, 356, 477, and 145 redds were counted for Sections 1 through 4 respectively. Maximum number of redds per section declined from 203 in Section 1 to 102, 122, and 46 in Sections 3, 4, and 5 respectively.

## Population Composition

Coded wire tagged fish comprised 21 \% of the total tagged carcasses based on the ratio of adipose fin clipped fish to total tagged carcasses (Table 3). Skeletons were not checked for adipose fin clips due to their advanced state of decomposition. However, it is likely that ratios calculated for tagged fish are representative for skeletons as well. The total contributions (tagged fish only) to the spawning population were $32 \%$ for natural males, $9 \%$ for CWT males, $47 \%$ for natural females, and $12 \%$ for CWT females (Figure 9). CWT verification and tag reading will be conducted at a later date therefore all CWT data presented here are preliminary.

Length frequency histograms of male and female fish (both natural and CWT) display bimodal peaks (Figures 10-13). The first peaks are likely grilse (age 1 and 2 fish) and the second peaks are likely adult (age 3, 4, and 5 year fish). Total grilse composition was $10 \%$ of the Tuolumne River escapement estimate. Breakpoints between grilse and adult were determined from basin wide fork length data. Breakpoints used were $<60 \mathrm{~cm}$ for natural females, $<62$ for adipose fin clipped females, 68 cm for natural males and 68 cm for adipose fin clipped males. Further breakdown of grilse is presented in Table 7.

## Sample Collection

Scales, otolith, and DNA samples were collected from both natural and adipose fin clipped fish throughout the survey period and survey area (Tables 8,9 and 10). Distribution of sampling is intended to best represent the spawning population over time, space, and origin. Scale and otolith samples will be utilized in the CDFG age determination program and for subsequent cohort analysis of San Joaquin River Basin Chinook salmon populations. One-hundred DNA samples were collected and delivered to the CDFG Salmonid Tissue Archives.

## Egg Production Estimate

An estimate of egg production by the 2003 fall run Chinook salmon is done using the relationship of fork length to fecundity. The relationship was developed using 48 San Joaquin fall run Chinook females ranging from fork length 62.5 to 94.0 cm (Loudermilk et al. 1990). The number of eggs was calculated for natural females ( $\mathrm{n}=277$, average $\mathrm{FL}=77.1$ ) and CWT females ( $\mathrm{n}=71$, average $\mathrm{FL}=78.3$ ) and then expanded to the entire estimate. Natural females made up $47 \%$ of the 2003 estimate and produced
approximately $6,194,673$ eggs. Adipose fin clipped females (12\%) produced approximately 1,628,784 eggs.

## Tuolumne River Flows

Tuolumne River flows at the La Grange guage ranged from approximately 210cfs to 470cfs during the 2003 spawning season (Figure 14). To attract fish into the Tuolumne from the San Joaquin River and improve spawning habitat a pulse flow was initiated on 15 October 2003. Flow increased to approximately 470cfs on 16 October 2003 and ramped down to 230cfs on 28 October 2003 and then decreased to about 210cfs for the remainder of the spawning season.

## Tuolumne River Temperature

Water temperatures are recorded in several locations throughout the spawning reach using data loggers placed and maintained by CDFG. Four sites are plotted in Figure 14.

## DISCUSSION

## Spawning Distribution

Redd counts are strongly affected by time of day, visibility, sunlight, wind rippling the water surface, redd superimposition, and other physical factors as well as the natural variability between observers. Furthermore, redd counts are conducted with a single pass as opposed to an intensive systematic approach beyond the scope of this study. In the primary spawning riffles of Section 1 the problem of redd superimposition is acute and leads to undercounting. On the other hand, redds in Section 2, 3, and 4 are easily delineated as clean patches of freshly worked gravel among patches of darker undisturbed gravel. In these sections redd counts are accurate indicators of spawning density. For these reasons, the disparity between spawning density in Section 1 versus Sections 2, 3, and 4 is likely greater than displayed in Figures 10 and 11.

## Population Estimate

The 2003 tag recovery rate of $55.3 \%$ is lower than the $64.4 \%$ in 2002 and the $61.3 \%$ reported in 2001, which are high tag recovery rates, but still higher compared to the recovery rate of $41.7 \%$ encountered in 2000 (Vasques 2001). The difference in recovery rates is likely a function of the difference in stream flow between 2000, (over 300cfs) and 2001-2003, (under 200cfs). Stream flow dynamics affects the likelihood of collecting carcasses in that it effects both how carcasses are distributed in the system and the
effectiveness in recovering carcasses by field crews. During the lower flows encountered during the 2002 and 2003 surveys carcasses were easily visible and the lower flows allowed for collection in specific locations which were too deep or too swift to survey in 2000. Furthermore, the banks of riffles were walked in an effort to collect carcasses that could not be seen or collected during the initial float through the riffle and subsequent pool. During 2000 bank efforts were not nearly so extensive. The Tuolumne River escapement estimate for 2003 of 2,163 salmon is the lowest since the 1996 estimate of 4,550 salmon.

## Population Composition

Coded wire tagged fish comprised 21 \% of the total tagged carcasses based on the ratio of adipose fin clipped fish to total tagged carcasses (Table 3). Skeletons were not checked for adipose fin clips due to their advanced state of decomposition. However, it is likely that ratios calculated for tagged fish are representative for skeletons as well. The total contributions (tagged fish only) to the spawning population were $32 \%$ for natural males, $9 \%$ for adipose fin clipped males, $47 \%$ for natural females, and $12 \%$ for adipose fin clipped females (Figure 12). CWT verification and tag reading will be conducted at a later date therefore all CWT data presented here are preliminary.

Length frequency histograms of male and female fish (both natural and CWT) display bimodal peaks (Figures $10,11,12$ and 13). The first peaks are likely grilse (age 1 and 2 fish) and the second peaks are likely adult (age 3, 4, and 5 year fish). Total grilse composition was $10 \%$ of the Tuolumne River escapement estimate. Breakpoints between grilse and adult were determined from basin wide fork length data and applied to Tuolumne River fork length data to determine grilse composition. Breakpoints used were $<66 \mathrm{~cm}$ for natural females, $<68 \mathrm{~cm}$ for adipose fin clipped females, $<72 \mathrm{~cm}$ for natural males and $<67 \mathrm{~cm}$ for adipose fin clipped males. Further breakdown of grilse is presented in Table 7.

## Tuolumne River Flows

Low dissolved oxygen levels in the San Joaquin River are believed to be a barrier for fall-run salmon migrating up the San Joaquin stem to spawn in the Merced, Tuolumne and Stanislaus Rivers. A fall pulse flow regime has been developed to lower river temperatures and elevate levels of dissolved oxygen in the San Joaquin River in order to attract salmon and prevent straying. Live salmon counts on the Tuolumne River peaked in week 7 and coincided with the end of the elevated dissolved oxygen levels, derived from the fall pulse flows, in the San Joaquin River. The flow, temperatures, observed live fish and redds are presented in Figure 16.

## Tuolumne River Temperatures

Temperatures in the upper sections (Section 1 and 2) down to Tuolumne River State Recreation Area (TRSRA)(RM 41.7) remained below the maximum thermal limit of $13.3^{\circ} \mathrm{C}$ for most all of the spawning season except for a few days in early October. This temperature is considered to be the upper thermal limit for successful egg incubation (Myrick and Cech 1998). River temperatures at Hickman Bridge fell below the $13.3^{\circ} \mathrm{C}$ level in the beginning of November and coincided with the first redd observations in week 5 of the survey. Temperatures remained below the benchmark $13.3^{\circ} \mathrm{C}$ for about a week and the decreased further which coincided with the peak of redd observations in weeks 8 and 9 . A slight increase in temperature seen at the Hickman Bridge location also saw slight decrease in live fish observations.

Table 1. Tuolumne River riffle identification cross-reference, 2003 to 2002.

| Section 1 |  | Section 2 |  | Section 3 |  | Section 4 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| New ID | Old ID | New ID | Old ID | New ID | Old ID | New ID | Old ID |
| 1a | A1A | F1 | F1 | K1 | K1 | S1 | S1 |
| A1n | A1 | F2 | F2 | K2 | K2 | S2 | S2 |
| A1s | A1 | F3 | F3 | L1 | L1 | S3 | S3 |
| A2 | A2 | G1N | G1 | L2 | L2 | T1 | T2 |
| B1 | B1 | G1S | G1 | L3 | L3 | T2 | T3 |
| B2 | B2 | G2 | G2 | M1 | None | T3 | T4 |
| B3 | B3 | G3 | G3 | M2 | None | T4 | T5 |
| C1 | C1 | G4 | G4 | N1 | None | T5 | None |
| C2 | C1 | H1 | H1 | N2 | None | U1 | U1 |
| C3 | C3 | H2 | H2 | N3 | N3 | U2 | U2 |
| D1 | D1 | H3N | H3 | N4 | N4 | U3 | U3 |
| D2 | D2 | H3S | H4 | O1 | O1 | V1 | V1 |
| D3 | D3 | H4 | H5 | O2 | O3 | V2 | V2 |
| D4 | D4 | H5 | H6 | O3 | None | V3 | V3 |
| D5 | D5 | H6 | H7 | O4 | O4 | V4 | V4 |
| E1 | E1 | I1 | I1 | O5 | O5 | W1 | W1 |
|  |  | I2 | I2 | P1 | P1 | W2 | W2 |
|  |  | I3 | I3 | P2 | P2 | W3 | W3 |
|  |  | J1 | J1 | P3 | P3 | X1 | X1 |
|  |  | J2 | J2 | P4 | P4 | X2 | X2 |
|  |  | J3 | J3 | Q1 | Q1 |  |  |
|  |  | J4 | J4 | Q2 | Q2 |  |  |
|  |  | J5 | J5 | Q3 | Q3 |  |  |
|  |  |  |  | R1 | R1 |  |  |
|  |  |  |  | R2 | R2 |  |  |
|  |  |  |  | R3 | R3 |  |  |

Table 2. Total weekly counts of live fish, redds, and carcasses.

| Week | Live | Redds | Carcasses |
| :---: | :---: | :---: | :---: |
| $\mathbf{1}$ | 2 | 0 | 1 |
| $\mathbf{2}$ | 38 | 0 | 2 |
| $\mathbf{3}$ | 66 | 0 | 1 |
| $\mathbf{4}$ | 203 | 3 | 2 |
| $\mathbf{5}$ | 395 | 99 | 17 |
| $\mathbf{6}$ | 343 | 180 | 100 |
| $\mathbf{7}$ | 462 | 217 | 164 |
| $\mathbf{8}$ | 463 | 349 | 367 |
| $\mathbf{9}$ | 342 | 255 | 364 |
| $\mathbf{1 0}$ | 196 | 149 | 237 |
| $\mathbf{1 2}$ | 151 | 215 | 117 |
| $\mathbf{1 3}$ | 89 | 131 | 87 |
| $\mathbf{1 4}$ | 52 | 24 | 28 |
| $\mathbf{1 5}$ | 6 | 4 | 12 |
| Total | 2810 | 1 | 9 |

${ }^{\text {a }}$ Carcasses includes all tagged carcasses and skeletons but does not include recoveries.

Table 3. Weekly totals.

| Week | Total Tagged | Skeletons | Fresh <br> Recoveries | Total <br> Counted |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 0 | 1 | 0 | 1 | Fresh Tagged | CWT's |
| 1 | 1 | 1 | 0 | 2 | 0 | 0 |
| 2 | 0 | 1 | 1 | 2 | 0 | 0 |
| 3 | 1 | 1 | 0 | 2 | 1 | 0 |
| 4 | 16 | 1 | 0 | 17 | 15 | 0 |
| 5 | 52 | 48 | 4 | 104 | 51 | 1 |
| 6 | 78 | 85 | 19 | 182 | 67 | 13 |
| 7 | 157 | 210 | 42 | 409 | 129 | 22 |
| 8 | 134 | 230 | 93 | 457 | 101 | 42 |
| 9 | 80 | 157 | 52 | 289 | 62 | 33 |
| 10 | 34 | 83 | 26 | 143 | 28 | 10 |
| 11 | 21 | 66 | 24 | 111 | 19 | 1 |
| 12 | 10 | 18 | 2 | 30 | 10 | 0 |
| 14 | 0 | 12 | 2 | 14 | 0 | 1 |
| 15 | 0 | 9 | $\mathbf{9 2 3}$ | $\mathbf{2 6 6}$ | $\mathbf{1 7 7 3}$ | $\mathbf{4 8 4}$ |
| Total | $\mathbf{5 8 4}$ |  |  |  | 0 | $\mathbf{1 2 3}$ |

[^0]Table 4. Distribution of fresh tagged fish, tag week versus recovery week.

| Recovery <br> Week | Tag Week of Recovered Tags |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ | $\mathbf{5}$ | $\mathbf{6}$ | $\mathbf{7}$ | $\mathbf{8}$ | $\mathbf{9}$ | $\mathbf{1 0}$ | $\mathbf{1 1}$ | $\mathbf{1 2}$ | $\mathbf{1 3}$ |
| $\mathbf{2}$ | 0 |  |  |  |  |  |  |  |  |  |  |  |  |
| $\mathbf{3}$ | 0 | 1 |  |  |  |  |  |  |  |  |  |  |  |
| $\mathbf{4}$ | 0 | 0 | 0 |  |  |  |  |  |  |  |  |  |  |
| $\mathbf{5}$ | 0 | 0 | 0 | 0 |  |  |  |  |  |  |  |  |  |
| $\mathbf{6}$ | 0 | 0 | 0 | 0 | 4 |  |  |  |  |  |  |  |  |
| $\mathbf{7}$ | 0 | 0 | 0 | 0 | 0 | 19 |  |  |  |  |  |  |  |
| $\mathbf{8}$ | 0 | 0 | 0 | 0 | 0 | 4 | 38 |  |  |  |  |  |  |
| $\mathbf{9}$ | 0 | 0 | 0 | 0 | 0 | 2 | 8 | 83 |  |  |  |  |  |
| $\mathbf{1 0}$ | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 49 |  |  |  |  |
| $\mathbf{1 1}$ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 8 | 16 |  |  |  |
| $\mathbf{1 2}$ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | 8 | 13 |  |  |
| $\mathbf{1 3}$ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 |  |
| $\mathbf{1 4}$ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 |
| $\mathbf{1 5}$ | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
| Fresh <br> Recoveries | 0 | 1 | 0 | 0 | 4 | 25 | 47 | 87 | 60 | 25 | 13 | 3 | 1 |
| Fresh Tagged <br> Carcasses | 0 | 1 | 0 | 1 | 15 | 51 | 67 | 129 | 101 | 62 | 28 | 19 | 10 |
| Percent <br> Recovery | $\mathbf{0 . 0}$ | $\mathbf{1 0 0 . 0}$ | $\mathbf{0 . 0}$ | $\mathbf{0 . 0}$ | $\mathbf{2 6 . 7}$ | $\mathbf{4 9 . 0}$ | $\mathbf{7 0 . 1}$ | $\mathbf{6 7 . 4}$ | $\mathbf{5 9 . 4}$ | $\mathbf{4 0 . 3}$ | $\mathbf{4 6 . 4}$ | $\mathbf{1 5 . 8}$ | $\mathbf{1 0 . 0}$ |

Table 5. Weekly Shaefer and Jolly-Seber estimates.

| Recovery Week | Number of Tags recovered | Total Carcasses Handled | Fresh Fish |  | All Fish |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Shaefer Estimate | Jolly-Seber Estimate | Jolly-Seber Estimate |
| 1 | 0 | 6 | 0 | 32 | 32 |
| 2 | 1 | 24 | 33 | 159 | 164 |
| 3 | 4 | 104 | 339 | 319 | 315 |
| 4 | 19 | 182 | 304 | 504 | 534 |
| 5 | 42 | 409 | 478 | 364 | 349 |
| 6 | 93 | 457 | 580 | 402 | 372 |
| 7 | 52 | 289 | 421 | 198 | 171 |
| 8 | 26 | 143 | 281 | 60 | 86 |
| 9 | 24 | 111 | 226 | 155 | 128 |
| 10 | 2 | 30 | 122 | -4 | 5 |
| 11 | 2 | 14 | 114 | 6 | 6 |
| 12 | 1 | 10 | 63 | 0 | 0 |
|  | Total Estimate |  | $\begin{gathered} \text { Shaefer (Fresh) } \\ 2,961 \end{gathered}$ | Jolly-Seber (Fresh) 2,195 | $\begin{gathered} \text { Jolly-Seber (All) } \\ 2,163 \end{gathered}$ |

Table 6. Maximum redd count for each riffle over the course of the escapement survey by section.

| Section 1 |  | Section 2 |  | Section 3 |  | Section 4 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Riffle | Maximum \# of Redds | Riffle | Maximum \# of Redds | Riffle | Maximum \# of Redds | Riffle | Maximum \# of Redds |
| 1a | 1 | F1 | 10 | K1 | 8 | S1 | 5 |
| A1 | 3 | F2 | 9 | K2 | 11 | S2 | 3 |
| A1n | 5 | F3 | 5 | L1 | 6 | S3 | 5 |
| A1s | 6 | G1N | 1 | L2 | 6 | T1 | 1 |
| A2 | 1 | G1S | 7 | L3 | 4 | T2 | 4 |
| B1 | 28 | G2 | 6 | M1 | 1 | T3 | 2 |
| B2 | 20 | G3 | 4 | M2 | 2 | T4 | 4 |
| B3 | 18 | G4 | 2 | N1 | 3 | T5 | 4 |
| C1 | 16 | G4p | 1 | N2 | 5 | U1 | 5 |
| C2 | 0 | H1 | 3 | N3 | 1 | U2 | 2 |
| C3 | 28 | H2 | 7 | N4 | 6 | U3 | 0 |
| D1 | 12 | H3N | 1 | O1 | 5 | V1 | 4 |
| D2 | 22 | H3S | 7 | O2 | 4 | V2 | 0 |
| D3 | 16 | H4 | 2 | O3 | 6 | V3 | 1 |
| D4 | 13 | H5 | 4 | O4 | 1 | V4 | 2 |
| D5 | 6 | H6 | 4 | O5 | 5 | W1 | 0 |
| E1 | 8 | I1 | 3 | P1 | 0 | W2 | 4 |
|  |  | I2 | 3 | P2 | 7 | W3 | 0 |
|  |  | I3 | 2 | P3 | 7 | X1 | 0 |
|  |  | J1 | 2 | P4 | 2 | X2 | 0 |
|  |  | J2 | 5 | Q1 | 10 |  |  |
|  |  | J3 | 4 | Q2 | 5 |  |  |
|  |  | J4 | 8 | Q3 | 6 |  |  |
|  |  | J5 | 2 | R1 | 4 |  |  |
|  |  |  |  | R2 | 2 |  |  |
|  |  |  |  | R3 | 5 |  |  |
| Subtotal | 203 |  | 102 |  | 122 |  | 46 |
| Total Redds | 473 |  |  |  |  |  |  |

Table 7. Grilse composition of Chinook salmon.

|  | Male | Female | Male (n=235) |  | Female (n=349) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Natural | Adclip | Natural |  |
| Grilse | $\mathbf{7 \%}$ <br> $(\mathrm{n}=40)$ | $\mathbf{3 \%}$ <br> $(\mathrm{n}=19)$ | $5 \%$ <br> $(\mathrm{n}=12)$ | $\mathbf{1 2 \%}$ <br> $(\mathrm{n}=28)$ | $\mathbf{1 \%}$ <br> $(\mathrm{n}=4)$ | $\mathbf{4 \%}$ <br> $(\mathrm{n}=15)$ |
| Adult | $\mathbf{3 3 \%}$ <br> $(\mathrm{n}=195)$ | $\mathbf{5 7 \%}$ <br> $(\mathrm{n}=330)$ | $\mathbf{1 7 \%}$ <br> $(\mathrm{n}=39)$ | $\mathbf{6 6 \%}$ <br> $(\mathrm{n}=156)$ | $\mathbf{1 9 \%}$ <br> $(\mathrm{n}=67)$ | $75 \%$ <br> $(\mathrm{n}=263)$ |

Table 8. Distribution of scale samples collected by section and week for natural and adipose fin clipped salmon.

| Week | Section |  |  |  | Weekly Total |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ |  |
| $\mathbf{1}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{2}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{3}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{4}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{5}$ | 4 | $0(1)$ | 0 | 0 | 5 |
| $\mathbf{6}$ | $12(3)$ | 1 | 0 | 0 | 16 |
| $\mathbf{7}$ | $12(7)$ | $2(1)$ | $1(2)$ | 0 | 25 |
| $\mathbf{8}$ | $28(12)$ | 5 | $2(2)$ | 2 | 51 |
| $\mathbf{9}$ | $24(7)$ | $4(3)$ | $2(2)$ | 0 | 42 |
| $\mathbf{1 0}$ | $14(3)$ | 5 | 2 | 2 | 26 |
| $\mathbf{1 1}$ | 7 | 1 | 0 | 2 | 10 |
| $\mathbf{1 2}$ | 5 | 0 | 1 | 1 | 7 |
| $\mathbf{1 3}$ | 1 | 1 | 0 | 1 | 3 |
| Section Totals | 139 | 24 | 14 | 8 | 185 |

Parenthesis indicate number of samples from adipose fin-clipped carcasses.

Table 9. Distribution of heads collected from Chinook salmon.

| Week | Section |  |  |  | Weekly Total |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ |  |
| $\mathbf{1}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{2}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{3}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{4}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{5}$ | 0 | 1 | 0 | 0 | 1 |
| $\mathbf{6}$ | 13 | 0 | 0 | 0 | 13 |
| $\mathbf{7}$ | 19 | 1 | 2 | 0 | 22 |
| $\mathbf{8}$ | 36 | 2 | 4 | 0 | 42 |
| $\mathbf{9}$ | 22 | 6 | 5 | 0 | 33 |
| $\mathbf{1 0}$ | 9 | 0 | 1 | 0 | 10 |
| $\mathbf{1 1}$ | 1 | 0 | 0 | 0 | 1 |
| $\mathbf{1 2}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{1 3}$ | 0 | 0 | 0 | 0 | 1 |
| Section Totals | 101 | 10 | 12 | 0 | 123 |

Heads were taken only from adipose fin-clipped carcasses.

Table 10. Distribution of DNA samples collected from natural and adipose fin clipped salmon.

| Week | Section |  |  |  | Weekly Total |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ |  |
| $\mathbf{1}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{2}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{3}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{4}$ | 0 | 0 | 0 | 0 | 0 |
| $\mathbf{5}$ | 0 | 0 | 1 | 0 | 1 |
| $\mathbf{6}$ | $5(1)$ | 0 | 0 | 0 | 6 |
| $\mathbf{7}$ | $11(5)$ | 3 | $2(1)$ | 0 | 22 |
| $\mathbf{8}$ | $12(4)$ | 3 | 2 | 1 | 22 |
| $\mathbf{9}$ | 9 | 2 | 1 | 1 | 13 |
| $\mathbf{1 0}$ | 3 | 9 | 3 | 4 | 19 |
| $\mathbf{1 1}$ | 11 | 2 | 0 | 3 | 16 |
| $\mathbf{1 2}$ | 1 | 0 | 0 | 0 | 1 |
| $\mathbf{1 3}$ | 0 | 0 | 0 | 0 | 0 |
| Section Totals | 62 | 19 | 10 | 9 | 100 |

Parenthesis indicate number of samples from adipose fin-clipped carcasses.



Figure 2. Fresh carcass indicated by clear eye.


Figure 3. Fungus covered skeleton.


Figure 4. Two skeletons showing varied degrees of decomposition and a fresh carcass.


Figure 5. Live fish observation, redd, and total carcass weekly counts. Total carcasses includes all tagged carcasses and skeletons.


Figure 6. Weekly cumulative Schaeffer and Jolly-Seber escapement estimates.


Figure 7. Total number of redds counted per section.


Figure 8. Total redds observed by riffle section. Each letter represents one river mile.


Figure 9. Contribution of natural female, adipose clipped female, natural male, and adipose fin clipped male to the 2003 Tuolumne River escapement.


Figure 10. Length frequency histogram of natural male Chinook salmon.


Figure 11. Length frequency histogram of adipose fin clipped male Chinook salmon.


Figure 12. Length frequency histogram of natural female Chinook salmon.


Figure 13. Length frequency histogram of adipose fin clipped female Chinook salmon.


Figure 14. Average daily flow in the Tuolumne River (cubic feet per second) at the Modesto, and La Grange gauges. Preliminary data obtained from California Data Exchange Center (CDEC) website.


Figure 15. Average daily temperature ( ${ }^{\circ} \mathrm{C}$ ) in the Tuolumne River at Hickman Bridge, RM 37.1, Turlock State Recreation Area, RM 41.8, Riffle D2, RM 48.9, and Riffle 1A, RM 53.0. Temperatures where obtained from thermograph data collected by CDFG.


Figure 16. Weekly live salmon counts for the Tuolumne River escapement survey. Flow (cfs) at La Grange guage, temperatures from CDFG monitoring sites, maximum thermal limit.

## References

Fry, D.H. 1961. King Salmon Spawning Stocks of The California Central Valley, 1949-1059. Calif. Fish and Game 47(1); 55-71.

Heyne, T. 2000. Tuolumne River Chinook Salmon Spawning Escapement Survey Federal Energy Regulatory Commission Annual Report FERC Project \#2299, Report 2001-2.

Heyne, T. 1998 Tuolumne River Chinook Salmon Spawning Escapement Survey Federal Energy Regulatory Commission Annual Report FERC Project \#2299, Report 1999-2.

Kano, R. M. 1999. Chinook salmon spawner stocks in California’s Central Valley, 1995. Inland Fisheries Admin. Report No. 99-7.

Law, P.M.W. 1994. Simulation study of salmon carcass survey capture-recapture methods. Calif. Fish and Game 80(1); 14-28.

Loudermilk, W., Neillands, W., Fjelstad, M., Chadwick, C., and Shiba, S. 1990. Annual Performance Report. Inland and Anadromous Sport Fish Management and Research. Project F-51-R-1. Job 2. 7pp.

Myrick, C. A. and Cech, J. J. Jr. 1998. Bay-Delta Modeling Forum Technical Publication 01-1. Temperature Effects on Chinook Salmon and Steelhead: A Review Focusing on California's Central Valley Populations. 57pp.

Niellands, W. George, Shiba, S., Baumgartner, S., Kleinfelter, J. 1993. Annual Performance Report. Inland and Anadromous Sport Fish Management and Research. Project F-51-R-1. Job 2. 34pp.

Ricker, W. E. 1975. Computation and interpretation of biological statistics of fish populations. Dept. of the Env. Fisheries and Marine Service, Bull., 191, 382pp.

Seber, G. A. F., 1973, Estimation of animal abundance and related parameters, Griffin, London, 506pp

Schaefer, M. B. 1951. Estimation of the size of animal populations by marking experiments. U.S. Fish and Wildlife Service Bull., 52:189-203.

Vasques, J. 2001. 2000 Tuolumne River Chinook Salmon Spawning Escapement Survey. Federal Energy Regulatory Commission Annual Report FERC Project \#2299, Report 2002-2.

Yoshiyama, R.M., E.R. Gerstung, F.W. Fisher, and P.B. Moyle. 2000. Chinook salmon in the California Central Valley: an assessment. Fisheries 25(2):6-19.


[^0]:    ${ }^{1}$ Includes only fish that were deemed fresh when tagged.
    ${ }^{2}$ Includes total tagged, skeletons, and fresh recoveries.

